HUMACYTE®

TISSUE-ENGINEERED HUMAN ACELLULAR BLOOD VESSELS FOR CORONARY ARTERY BYPASS GRAFTING

Adam R. Williams, MD¹, Joseph Nellis, MD¹, Zachary K. Wegermann, MD¹, Sharon L. McCartney, MD¹, Mihai V. Podgoreanu, MD¹, Sachin Mehta, MD¹, Kyha D. Williams, DVM¹, Melissa A. Daubert, MD¹, Robert D. Kirkton, PhD², Kevin M. Nash, PhD², Laura E. Niklason, MD, PhD², Jeffery H. Lawson, MD, PhD¹, Alan P. Kypson, MD^{2,3}

¹Duke University, Durham, NC; ²Humacyte,Inc, Durham, NC; ³REX Hospital and the University of North Carolina, Raleigh, NC

INTRODUCTION

Coronary artery disease (CAD) is the narrowing of the major blood vessels of the heart with symptoms ranging from angina to myocardial infarction and death. Treatment includes medical therapy and procedures such as percutaneous coronary interventions (PCI) or coronary artery bypass grafting (CABG). CABG is among the most common surgeries performed in the US (~400,000 surgeries per year)¹ and has been shown to improve quality of life as well as survival. The primary conduits are left internal mammary artery (LIMA) and saphenous vein, which is used in 80-90% of CABG in the US. SVG patency at 1 year is reported to be as low as 75%, primarily due to thrombosis or neo-intimal hyperplasia.² Additionally, SVG harvest can result in surgical wound infection at the harvest site potentially leading to prolonged hospital stay, need for revascularization and limb-loss.³ Therefore, an unmet need exists for an advanced, readily available CABG conduit for the treatment of CAD.

The Human Acellular Vessel (HAV), developed at Humacyte, Inc., is an investigational tissue-engineered blood vessel consisting of human extracellular matrix (ECM) proteins. The HAV is created by culturing human vascular cells within a biodegradable scaffold under biochemical and biomechanical stimulation. The resulting tissue is then decellularized to yield a mechanically robust and non-immunogenic HAV (Fig 1).⁴ The HAV has been shown to be remodeled by the recipient's own cells to closely resemble native vasculature.⁵ In this study, we used a small diameter (3.5mm) HAV as a conduit for CABG in a baboon surgical model and investigated the in-vivo remodeling of the human HAV in the baboon for up to 6 months.

SMALL DIAMETER HUMAN ACELLULAR **VESSELS (HAVS)**



▲ Figure 1. Schematic of HAV production

EXPERIMENTAL DESIGN

Small diameter (3.5mm) HAVs were produced at Humacyte, Inc. by scaling down our investigational 6mm HAV platform. Healthy adult male adult baboons (Papio anubis / Papio cynocephalus, n=7, 26.8 – 37.0kg) were acquired from Texas Biomedical (San Antonio, TX). Non-immunosuppressed baboons were surgically implanted with a 3.5mm HAV as a CABG conduit under cardiopulmonary bypass using standard surgical techniques. The bypassed native coronary was ligated proximal to the HAV anastomosis and flow through the HAV was confirmed by transit time flowmetry (TTFM). Baboons were given aspirin (3.5mg/kg) and clopidogrel (0.2-0.5 mg/kg) daily throughout the study.

▼Figure 2. Experimental Timeline

Imp Da	lant y 0 <u>1-2</u>	Week	<u>1 Mo</u>	Explant <u>6 Months</u>		
TT	FM Ca	ath	СТА	СТА	Cath	Histology & CTA



Aorta-LAD Bypass Aorta-RCA

IN VIVO LONGITUDINAL EVALUATION OF IMPLANTED HAV CABG CONDUITS





Figure 6. Histological evaluation of host remodeling. HAVs explanted at 6 Months from animals Bb5, Bb6 were stained by H&E (A, B), by Masson's Trichrome (C), for α-smooth muscle actin (αSMA, D, E) and von Willebrand Factor (vWF, D, E). Formation of a neo-adventitia and infiltration of host αSMA⁺ cells were observed throughout explanted HAVs. The presence of endothelial cells expressing vWF (white triangles, E) on the lumen were observed in 6-Month explants.



DEVELOPMENT OF A BABOON SURGICAL MODEL OF CABG

Table 1. Outcomes of 3.5mm HAV CABG Conduits in Baboons

	Animal	Weight (kg)	Configuration	HAV Length (cm)	TTFM (mL/min)	Implant Duration	Outcome
	Bb 1	30.2	Aorta-LAD	10.0	5	2 Weeks	Occluded
	Bb 2	30.0	Aorta-LAD	7.3	20	9 Weeks	Occluded
nterior nding nary ary	Bb 3	28.1	Aorta-RCA	7.5	17	0 Days	(Surgical Complication)
ass	Bb 4	26.8	Aorta-RCA	3.1	15	6 Weeks	Occluded
	Bb 5	31.5	Aorta-RCA	3.2	34	27 Weeks	Patent
	Bb 6	30.5	Aorta-RCA	3.5	19	23 Weeks	Patent
	Bb 7	37.0	Aorta-RCA	2.0	25	17 Weeks*	Patent*
	*In-life pl	hase ongoin	ng, planned duration	n: 6 Months			

✓ Figure 3. CABG Surgical Model Development. Baboon (Bb) 1 and Bb2 were implanted with an aorta-LAD HAV bypass (A, B) which was occluded shortly post-op due to size-mismatch. Animals Bb3-7 were implanted with an aorta-RCA HAV bypass (C, D) which resulted in improved success and patency out to 6 Months.

> ◄ Figure 4. Computed Tomography Angiography (CTA) 3D Reconstruction evaluation of HAV CABG conduits at 1 Month (A, B), 3 Months (C, D) and 6 Months (E, F) post-implant. Noninvasive imaging of implanted baboons showed no dilation significant stenosis of or aorta-RCA HAV conduits through 6 Months.

Figure 5. Following Surgical Implantation of Aorta-RCA HAV conduits on Day 0 (A), HAVs were evaluated by left heart catheterization at 1-2 Weeks (B) and 6 Months (C) prior to HAV explant (D). Catheter angiography imaging at 6 Months showed patent HAV CABG conduits in animals Bb5 and Bb6. Explanted HAVs were found to be well incorporated to host tissue.

HOST REMODELING OF HAV CABG CONDUITS

BABOON CORONARY ANATOMY VARIABILITY



- CABG with sustained patency at 6 months.

- Cardiol. (2019).
- vessels after human implantation. Sci Transl Med. (2019).
- grafts. J Vasc Surg. (2012).
- Surg. (2013)

ACKNOWLEDGEMENTS

The authors would like to thank Derek Argotti CCP, Derek Sanderson Jr. CCP, Daniel Regan PA-C, and Christopher Rhew (CABG surgery), Lainie Parker, Kelly Shaw RVT, and Diego Zapata RVT, LAT (surgical support), Brittney Stone RT(R)(CT), and Keith Jastrow RT(R)(CT) (CTA support), Jeffrey Everitt DVM, and Yohannes G Asfaw DVM (Pathology).

These results are from a pre-clinical study. The authors employed at Humacyte, Inc. (RDK, KMN, LEN, JHL, and APK) own stock or stock options in Humacyte, Inc.

Duke Surgery Duke University School of Medicine

UNC REX HEALTHCARE II

To date there is only one previous report of the use of baboons in a surgical model of CABG.^{6,7} Zilla et al. successfully grafted autologous saphenous veins SVGs to healthy baboon (Papio ursinus) coronary arteries in an aorta-LAD configuration for up to 24 weeks (n=8 and n=18). During surgical implantation of our first two animals (Bb1, the baboon LAD was found to Bb2). be significantly smaller in diameter ($\sim 1.0 - 1.5$ mm), much thinner walled, and with poor distal runoff compared to humans (2.5 - 4.0 mm). The RCAs in Bbs 3-7 were considerably larger

(~2.0mm) and allowed for the long-term implantation of the 3.5 mm HAV in a clinically relevant configuration. The subsequent n=5 animals were implanted with a 3.5 mm HAV in an aorta-RCA configuration, which resulted in improved patency. Pre-operative CTA imaging revealed a varying baboon coronary anatomy with some animals exhibiting a larger RCA, LAD, or LCx artery. Additionally, n=2 animals not included in the study were found to have congenital malformations (dextrocardia and coronary fistulas) at pre-operative screening.

◄ Figure 7. Variability in Baboon Coronary Vascular Anatomy was observed by preoperative CTA imaging. Baboons were imaged before CABG to determine optimum distal target. Animals were shown to have right-dominant (A) and left-dominant coronaries with LAD- (B) and LCx-dominant (C) presentations.

SUMMARY

 Model development was challenging due to variability in coronary anatomy and HAV to host artery size-mismatch, but aorta-RCA configuration was most appropriate. • The 3.5 mm HAV was successfully used as a conduit in a baboon surgical model of

• The HAV repopulated with host endothelial, smooth muscle, and neoadventitial cells that remodeled the HAV conduit during the 6-month implantation.

REFERENCES

1. Alexander, JH., et al. Coronary-Artery Bypass Grafting. N Eng J Med. (2016). 2. Gaudino, M., et al. Mechanisms, consequences, and prevention of coronary graft failure. Circulation. (2017). 3. Caliskan, E., et al., Saphenous vein grafts in contemporary coronary artery bypass graft surgery Nat Rev

4. Dahl, S.L.M., et al., Readily available tissue-engineered vascular grafts. Sci Transl Med. (2011). 5. Kirkton, R.D., et al., Bioengineered human acellular vessels recellularize and evolve into living blood

6. Zilla, P., et al. Remodeling leads to distinctly more intimal hyperplasia in coronary than in infrainguinal vein

7. Moodley, L., et al. Protective constriction of coronary vein grafts with knitted nitinol. Eu J Cardio-Thorac

DISCLOSURE INFORMATION